

# Determination of cholesterol and cholesteryl esters in *Xenopus laevis* embryos through direct flow injection electrospray ionisation mass spectrometry

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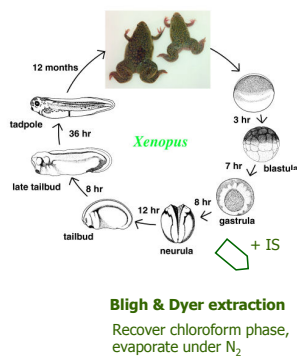
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## Introduction

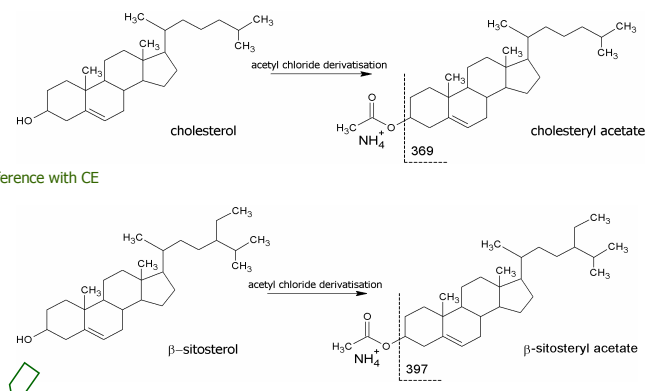
Cholesterol is an important structural component of animal cell membranes, especially in the liquid-ordered membrane microdomains, the so-called lipid rafts. Cholesteryl ester (CE) is the major transport and storage form of cholesterol. In the perspective of a biological question, a method for the quantification of cholesterol and CE is being developed. Cholesterol quantification is established in *Xenopus laevis* embryos in the early neurula stage together with the identification of the most important CE.

## Cholesterol quantification



### Experimental

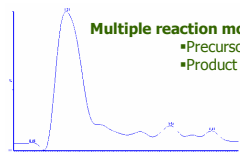
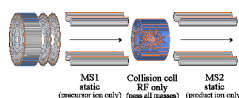
Acetylation  
 > No interference with CE



Agilent autosampler  
 Injection volume: 5 µl

Agilent 1100 pump  
 Flow rate: 10 µl/min  
 Eluent: MeOH = 100mM  
 Am. Acetate / CHCl<sub>3</sub> 3/1

### Direct flow injection electrospray MS/MS



### Multiple reaction monitoring

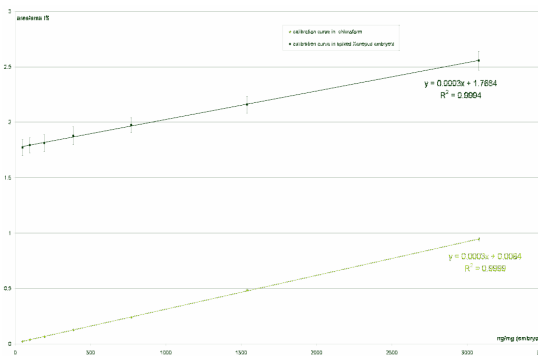
• Precursor ion:  $m/z$  446.4  
 • Product ion:  $m/z$  369.3

Data processing  
 Quanlynx®

## Results

Neutral free sterols are very poorly ionisable through electrospray ionisation (ESI). Cholesterol is hereby acetylated through a derivatisation step with acetyl chloride, resulting in an ionisable cholesteryl acetate (CE 2:0) by means of ammonium adduct formation. This is also compatible with the identification, and later on quantification, of CE. A fragment of  $m/z$  369.3 upon collision-induced fragmentation is generated. Quantification of cholesterol can thus be accomplished using a multiple reaction monitoring (MRM) transition of  $m/z$  446.4 to  $m/z$  369.3. This derivatisation also allows us to analyse both cholesterol and CE in a parallel way.

Quantification of cholesterol was performed using direct flow injection analysis of the *Xenopus* embryo extract. The method has a high throughput, a runtime of only 5 minutes is needed.  $\beta$ -Sitosterol, a typical plant sterol, was used as an internal standard. Blank *Xenopus* extracts did not show any interfering mass. As cholesterol is an endogenous compound, detection limit was approximated through spiking of  $\beta$ -sitosterol to *Xenopus* embryo extract. Using only 5 *Xenopus* embryos, a calibration curve of cholesterol (spiked to 5 WT *Xenopus* embryos) is established between 96 and 3000 ng/mg embryonic material.



Calibration curve for cholesterol concentration in pure chloroform and in spiked *Xenopus* extracts,  $n = 3$ . The intercept equals the cholesterol concentration in *Xenopus* embryo.

detection limit (ng/mg embryo)	repeatability (overall, n=50) (S-sitosterol)	LLOQ (ng/mg embryo)	precision (cholesterol)	accuracy	cholesterol content (µg/mg embryo)	variance analytical (n=9)	variance biological (n=3)
17.40	11.23%	96.15	4.42%	105.30%	5.54 ± 0.14	7.60%	10.59%

## Cholesteryl ester identification

In the perspective of a metabolic profiling method for cholesterol and its esters, a precursor ion scan method was developed. All precursor ions with the specific fragment of  $m/z$  369.3 are scanned in the range from  $m/z$  500 to 800. Cholesteryl 10-undecanoate is used as an internal standard to quantify the CE in a later stadium. Derivatisation did not interfere with CE identification. Most of the detected peaks can be addressed to common CE (see in figure).

## Conclusion

- Cholesterol can be quantified in a fast and reproducible way in *Xenopus* embryos. Notwithstanding the high concentration present in these embryos, a LLOQ of 96.15 ng/mg embryo is feasible.
- The mean cholesterol concentration in the early neurula stage is 5.54 ng/mg embryo, with a biological variance of 10.59%.
- Cholesteryl esters can be identified in the same samples. Quantification of these compounds will be established soon.

